## ANTAGONISTS OF PLATELET ACTIVATING FACTOR FROM SWIETENIA MAHOGANI (L,) JACQ

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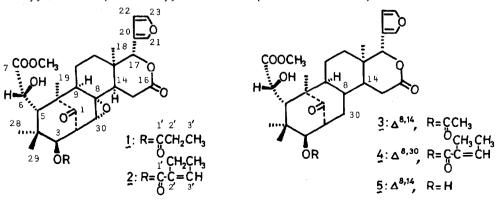
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Summary: The structures of swietemahonin A (1), E (2), and 3-acetylswietenolide (3), isolated as antagonists of platelet activating factor from the ether extract of <u>Swietenia mahogani</u> (L.) Jacq., were determined based on the 2-D NMR spectroscopy.

In our search for biologically significant substances from medicinal plants in Indonesia, we have isolated three new tetranortriterpenoids, swietemahonin A (1), E (2), and 3-acetyl-swietenolide (3), from the cotyledons of <u>Swietenia mahogani</u> (L.) Jacq.<sup>1)</sup> and found that they exhibit antagonistic effects on platelet activating factor (PAF).<sup>2)</sup> This paper describes the isolation and structure elucidation of these new tetranortriterpenoids.

The cotyledons (700 g) of <u>S. mahogani</u>, collected in Medan, North Sumatera-Indonesia, were pulverized and extracted with ether at room temperature and then with boiling MeOH. The ether extract yielded an oil and a solid precipitate. The oily fraction of the ether extract was roughly separated by silica gel column chromatography using benzene, benzene-CH<sub>2</sub>Cl<sub>2</sub>, CH<sub>2</sub>Cl<sub>2</sub>, and MeOH-CH<sub>2</sub>Cl<sub>2</sub>. The MeOH-CH<sub>2</sub>Cl<sub>2</sub> eluates were further separated by a combination of preparative TLC and HPLC to give swietemahonin A (1)(52 mg), E (2)(34 mg), and 3-acetylswietenolide (3)(56 mg), along with known swietenine (4)<sup>3</sup> and swietenolide (5).<sup>4</sup>

Swietemahonin A (1), colorless needles (from AcOEt-isopropyl ether), mp 174 - 174.5°C,  $\left[\alpha\right]_{D}$  -12.16 (CHCl<sub>3</sub>), has the molecular formula  $C_{30}H_{38}O_{10}$  (M<sup>+</sup> 558.2478, calcd. 558.2464) and its IR spectrum showed the presence of a hydroxyl (3500 cm<sup>-1</sup>), a lactone (1735 cm<sup>-1</sup>), an ester (1720 cm<sup>-1</sup>), a ketone (1700 cm<sup>-1</sup>), and a furan (1500 and 870 cm<sup>-1</sup>). The <sup>1</sup>H and <sup>13</sup>C NMR and



	swietenolide	( <u>3</u> ), Swiet	enine (4), a	and Swieteno	lide (5)
Compd 1 <sub>H</sub>	1	ş	3	4	5
2	3.54 dd (9.5, 2.5)	3.61 dd (9.5, 2.5)	3.15 ddd (10, 6, 2.5)	3.52 ddt (10, 8, 1.5)	3.04 ddd (10, 5.5, 2.5)
3	4,94 d (9,5)	4,84 d (9.5)	4.82 d (10)	4.64 d (10)	3.58 d (10)
5	3.27 br s	3.36 br s	3.22 br s	3.50 br s	3.25 br s
6	4.46 br s	4.46 brs	4.55 br s	4.56 br s	4.54 br s
15 a	2.79 dd {17, 7}	2.71 dd (17.5, 7.5)	3.47 dt (21, 3)	2.83 dd (18, 5)	3.46 dt (21, 2.5)
15 8	3.27 dd (17, 12.5)	3.16 dd (17.5, 12)	3.69 dt (21, 1.5)	2.76 dd (18, 2)	4.03 dt (21, 1)
18	1.05 s	1.04 s	1.03 5	0.97 s	0.99 s
19	1.34 s	1.35 s	1.42 s	1.45 s	1.40 s
28	1.11 s	1.12 s	1.09 s	1.12 s	1.00 s
29	0.88 s	0.92 s	0.80 s	0.89 s	0.88 s
30	3.20 d (2.5)	3.10 d (2.5)	2.15 m 2.81 dd (15.5, 2.5)	5.34 dt (8.0, 2)	2.01 dd (14.5, 5.5) 3.19 dd (14.5, 2.5)
2'	2.50 q (7.5)		2.15 s		(14.31 2.3)
2'-C <u>H</u> 3		1.94 s		1.82 s	
3'	1.24 t (7.5)	7.01 qq (6.0, 1.5)		6.87 qq (7.0, 1.5)	
з'-с <u>н</u> з		1.93 brd (6.0)		1.74 brd (7.0)	
соос <u>н</u> а	3.92 s	3.95 s	3.84 s	3.76 s	3.82 s

Table I. <sup>1</sup>H-NMR Spectral Data of Swietemahonin A (1), E (2), 3-Acetyl-. . . . . .

δ values in CDCl<sub>3</sub>. Values in parentheses are coupling constants (Hz).

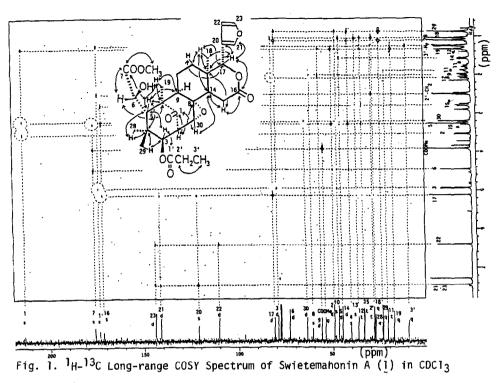
 $^{1}H^{-1}H$  and  $^{1}H^{-13}C$  COSY<sup>5</sup>) of 1 indicated the presence of a ketone ( $\delta_{C}$  213.51), a methyl ester  $(\delta_{H} 3.92; \delta_{C} 175.62 \text{ and } 53.50), \text{ a propionyl} (\delta_{H} 1.24 \text{ and } 2.50; \delta_{C} 9.24, 27.52, \text{ and } 173.14),$ a lactone  $(\delta_{\mu} 5.12; \delta_{C} 80.37 \text{ and } 171.20)$ , a furan  $(\delta_{\mu} 6.39, 7.43, \text{ and } 7.44; \delta_{C} 109.89, 140.69,$ 143.44, and 120.62), a hydroxymethine ( $\delta_{H}$  4.46;  $\delta_{C}$  72.40), an epoxide ( $\delta_{H}$  3.20;  $\delta_{C}$  60.17 and 63.23), four <u>tert</u>-methyl groups ( $\delta_{H}$  0.88, 1.05, 1.11, and 1.34;  $\delta_{C}$  22.72, 26.86, 23.10, and 17.01), and three quarternary sp<sup>3</sup> carbons ( $\delta_{C}$  35.91, 39.78, and 48.54)(Table I and III).

The above data led us to suppose that the structure of swietemahonin A might be ]. At this stage, we measured the  ${}^{1}H_{-}{}^{13}C$  long-range COSY<sup>5)</sup> of ] in order to confirm the assumed structure (1). As expected, the  $^{13}$ C-signals at  $\delta$  213.51 (C-1) and at  $\delta$  175.62 (C-7) showed long-range correlations with the <sup>1</sup>H-signals at  $\delta$  1.34 (19-H<sub>2</sub>), 3.20 (30-H), and 3.54 (2-H) and at  $\delta$  3.27 (5-H), 3.92 (COOCH<sub>3</sub>), and 4.46 (6-H), respectively (Fig. 1). In turn, the <sup>13</sup>C-signal at  $\delta$  173.14 (C-1') was correlated with the <sup>1</sup>H-signals at  $\delta$  1.24 (3'-H<sub>3</sub>), 2.50  $(2'-H_2)$ , and 4.94 (3-H), and the signal at  $\delta$  171.20 (C-16) with the <sup>1</sup>H-signals at  $\delta$  2.79, 3.27  $(15-H_2)$ , and 5.12 (17-H). Also some other significant long-range correlations are shown by arrows in formula in Fig. 1. Thus the planar structure of ] was proved.

The relative stereochemistry of 1 was determined on the basis of the coupling constants of each protons and the results of NOE experiments. Irradiation at the 29-H $_3$  and 18-H $_3$  caused the increase of signal intensity of the 28-, 2'-, 5-, 3-, and ester methyl-protons and the 12-, 14-, 17-, 22-, and 21-protons, respectively, and irradiation at the 28-H<sub>3</sub> and 19-H<sub>3</sub> enhanced the signal intensity of the 19- and 3-protons and the 28-, 9-, and 6-protons, respectively.

Therefore the structure of swietemahonin A was proved to be ].

Swietemahonin E ( $\underline{2}$ ), colorless needles (from AcOEt-isopropyl ether), mp 151 - 152°C,



 $[\alpha]_D$  -20.69 (CHCl<sub>3</sub>), has the molecular formula  $C_{32}H_{40}O_{10}$  (M<sup>+</sup> 584.2605, calcd. 584.2620) and it showed the IR spectrum very similar to that of 1. The <sup>1</sup>H and <sup>13</sup>C NMR spectra are almost identical with those of 1 except for the appearance of the signals due to the tigloyl residue instead of the signals due to the propionyl residue in 1. (Table I and III). From these comparisons

Table II. Inhibition of Rabbit Platelet Aggregation	Table	II.	Inhibition	of	Rabbit	Platelet	Aggregation
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Compound	Inhibition (%)
swietemahonin A (1)	97.4
swietemahonin E (2)	91.7
3-acetylswietenolide (3)	91.6
swietenine (4)	none
swietenolide (5)	35.2

Concentration of PAF (final 7.5 x  $10^{-8}$  M) Concentration of sample (final 100 µg/m1)

of  ${}^{1}$ H and  ${}^{13}$ C NMR data, it was concluded that in the structure of 2, the propionyl residue of 1 was replaced by the tigloyl residue at C-3.

3-Acetylswietenolide (3),  $C_{29}H_{36}O_{9}$ , colorless needles, mp 136 - 138°C,  $[\alpha]_D$  -17.76 (CHCl<sub>3</sub>), showed the <sup>1</sup>H-NMR spectrum closely similar to that of 5, except for the down-field shift of a proton doublet ( $\delta$  4.82) assignable to the 3-H (Table I). Acetylation of 3 gave a diacetate, which was identified as 3,6-diacetylswietenolide by comparing the spectral data with those reported in the literature.<sup>6</sup> Therefore 3 was considered to be the 3-acetyl derivative of 5.

The inhibitory effects on rabbit platelet aggregation<sup>7)</sup> of these tetranortriterpenoids are listed in Table II. It was found that swietemahonin A (1) showed the most strong anti-PAF activity.

Although various compounds such as ginkgolides, triazolobenzodiazepines, benzofuranoid neolignans, and furanoid lignans have been reported as antagonists of PAF,<sup>8)</sup> our present results

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Compd					
13 <sub>C</sub>	1	2	3	4 ~	5
1	213.51 (s)	213.70 (s)	217.59 (s)	216.53 (s)	219.80 (s)
2	48.68 (d)	49.01 (d)	47.86 (d)	48.95 (d)	49.99 (d)
2 3 4	78.44 (d)	79.13 (d)	79.94 (d)	78.41 (d)	78.50 (d)
4	39.78 (s)	40.05 (s)	38.72 (s)	39.04 (s)	39.66 (s)
5	46.26 (d)	46.16 (d)	45.22 (d)	45.47 (d)	44.00 (d)
6	72.40 (d)	72.39 (d)	73.33 (d)	72.87 (d)	73.57 (d)
6 7	175.62 (s)	72.39 (d) 175.85 (s)	175.45 (s)	175.97 (s)	175.83 (s)
8	60.17 (s)	60.30 (s)	128.52 (s)	138.28 (s)	129.05 (s)
9	55.45 (d)	55.08 (d)	53.59 (d)	57.56 (d)	52.96 (d)
10	48.54 (s)	48.65 (s)	53.23 (s)	50.39 (s)	53.99 (s)
11	20.38 (t)	20.63 (t)	18.96 (t)	21.28 (t)	18.74 (t)
12	32.91 (t)	32.59 (t)	29.76 (t)	34.64 (t)	29.08 (t)
13	35.91 (s)	35.65 (s)	38.08 (s)	36.73 (s)	37.81 (s)
14	44.30 (d)	43.52 (d)	131.46 (s)	45.09 (d)	130.75 (s)
15	32.65 (t)	32.50 (t)	33.59 (t)	29.57 (t)	33.15 (t)
16	171.20 (s)	170.78 (s)	169.44 (s)	168.45 (s)	171.43 (s)
17	80.37 (d)	80.86 (d)	80.92 (d)	76.71 (d)	80.51 (d)
20	120.62 (s)	120.93 (s)	120.72 (s)	121.38 (s)	120.81 (s)
21	140.69 (d)	140.63 (d)	141.12 (d)	140.54 (d)	141.06 (d)
22	109.89 (d)	109.82 (d)	109.70 (d)	109.24 (d)	109.80 (d)
23	143.44 (d)	143.48 (d)	143.09 (d)	143.20 (d)	142.88 (d)
30	63.23 (d)	63.04 (d)	33.89 (t)	123.66 (d)	33.80 (t)
18	26.86 (q)	27.06 (q)	18.41 (q)	21.28 (q)	17.91 (q)
19	17.01 (q)	17.07 (q)	17.53 (q)	16.53 (q)	17.91 (q)
28	23.10 (q)	23.24 (q)	23.33 (q)	22.80 (q)	23.22 (q)
29	22.72 (q)	23.24 (q)	23.08 (q)	23.05 (q)	23.63 (q)
1'	173.14 (s)	166.77 (s)	170.29 (s)	166.92 (s)	·
2'	27.52 (t)	127.88 (s)	21.14 (q)	127.77 (s)	
3'	9.24 (q)	139.54 (d)		139.02 (d)	
2'-CH2		12.34 (q)		11.75 (q)	
2'- <u>CH</u> 3'- <u>CH</u> 3		14.70 (q) 53.47 (q)		14.64 (a)	
COOCH3	53.50 (q)	53.47 (q)	53.17 (q)	53.28 (q)	53.23 (q)

Table III. <sup>13</sup>C-NMR Spectral Data of Swietemahonin A (1), E (2), 3-Acety1swietenolide (3), Swietenine (4), and Swietenolide (5)

 $\delta$  values in CDCl<sub>3</sub>. The multiplicities of carbon signals were determined by means of the off-resonance and are indicated as (s), (d), (t), and (q). Assignments were done by the use of  $^{1}\mathrm{H}\text{-}^{13}\mathrm{C}$  COSY and  $^{1}\mathrm{H}\text{-}^{13}\mathrm{C}$  long-range COSY.

provided the first examples of tetranortriterpenoids having antagonistic effects on PAF. Several tetranortriterpenoids were also isolated from <u>S. mahogani</u> and the investigation of their structures and anti-PAF activities is currently in progress.

## References and Notes

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- and platelet-poor plasma (PPP) was made at 1400 g for 10 min. Aggregation was measured by the changes in turbidity with Aggregometer, model RMA-31 (Rikadenki Kogyo, Japan) at 37°C. The aggregometer was adjusted in sensitivity to give light transmission values of 0 and 100 % for PRP and PPP, respectively. Inhibition of aggregation was assayed by comparing the change in the transmitted light in PAF-treated PRP with that in the sample-treated PRP.
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